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REVIEW ARTICLE

Modulation of tumor necrosis factor-related apoptosis-inducing ligand (TRAIL)-mediated apoptosis by *Helicobacter pylori* in immune pathogenesis of gastric mucosal damage



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Abstract *Helicobacter pylori* infection is associated with chronic gastritis, peptic ulcer, gastric carcinoma, and gastric mucosa-associated lymphoid tissue lymphomas. Apoptosis induced by microbial infections is implicated in the pathogenesis of *H. pylori* infection. Enhanced gastric epithelial cell apoptosis during *H. pylori* infection was suggested to play an important role in the pathogenesis of chronic gastritis and gastric pathology. In addition to directly triggering apoptosis, *H. pylori* induces sensitivity to tumor necrosis factor-related apoptosis-inducing ligand (TRAIL)-mediated apoptosis in gastric epithelial cells. Human gastric epithelial cells sensitized to *H. pylori* confer susceptibility to TRAIL-mediated apoptosis via modulation of death-receptor signaling. The induction of TRAIL sensitivity by *H. pylori* is dependent upon the activation of caspase-8 and its downstream pathway. *H. pylori* induces caspase-8 activation via enhanced assembly of the TRAIL death-inducing signaling complex through downregulation of cellular FLICE-inhibitory protein. Moreover, *H. pylori* infection induces infiltration of T lymphocytes and triggers inflammation to augment apoptosis. In *H. pylori* infection, significant increases in CCR6⁺ CD3⁺ T cell infiltration in the gastric mucosa was observed, and the CCR6 ligand, CCL20 chemokine, was selectively expressed in inflamed gastric tissues. These mechanisms initiate chemokine-mediated T lymphocyte trafficking into inflamed epithelium and induce mucosal injury during *Helicobacter* infection. This article will review recent findings on the interactions of *H. pylori* with host-epithelial signaling pathways and events involved in the initiation of gastric pathology, including gastric inflammation and mucosal damage.

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***Helicobacter pylori* infection and apoptosis in gastric epithelial cells**

Helicobacter pylori is a common human pathogen that infects over half of the world population and is associated with duodenal and peptic ulcer diseases. The clinical consequences range from asymptomatic gastritis to peptic ulceration and gastric malignancy.^{1,2} The outcome of the infection is determined by interactions among *H. pylori* virulence factors, host gastric mucosal factors, and the environment. However, the mechanisms by which host factors cause disease remain unclear. Apoptosis induced by microbial infections is implicated in the pathogenesis of *H. pylori* infection, and enhanced gastric epithelial cell apoptosis during *H. pylori* infection was suggested to play an important role in the pathogenesis of chronic gastritis and gastric pathology.^{3–7} Direct cytotoxicity, as well as inflammatory responses, occurs in gastric mucosa cells.^{5,8–10} *H. pylori* vacuolating (VacA) toxin induces gastric epithelial cell apoptosis, but cytotoxin-associated gene A (CagA) attenuates apoptosis in gastric epithelial cells. Bacterial virulent factors may contribute directly to apoptosis in epithelial cells, whereas the induction of sensitivity to death-receptor mediated apoptosis by *H. pylori* is independent of *H. pylori* virulent factors VacA and CagA,⁶ suggesting that immune factors, in addition to bacterial factors, are also important in determining the degree of gastric mucosa damage during *H. pylori* infection.

It was demonstrated that T helper type 1 (Th1) cells selectively increased during *H. pylori* infection.^{11–14} Th1 cytokines, such as interferon gamma (IFN- γ) and tumor necrosis factor alpha (TNF- α), can increase the release of proinflammatory cytokines, augmenting apoptosis induced by *H. pylori*.¹⁰ *H. pylori* infection could also induce gastric mucosa damage by increasing expression of Fas in gastric epithelial cells, leading to gastric epithelial cell apoptosis through Fas/Fas ligand (FasL) interaction with infiltrating T cells.^{9,15} These findings suggest a role for immune-mediated apoptosis of gastric epithelial cells during *H. pylori* infection. Additionally, *H. pylori* directly triggers cell death by cytotoxins after interacting with gastric epithelial cells.^{16,17} Furthermore, *H. pylori* translocates CagA into gastric epithelial cells by Type IV secretion, inducing intracellular protein phosphorylation and dysregulating signal transduction pathways within host cells.^{18–21} In addition to directly triggering apoptosis, it is possible that *H. pylori* induces sensitivity to apoptosis in gastric epithelial cells via modulation of apoptosis signaling.

***H. pylori* modulate tumor necrosis factor-related apoptosis-inducing ligand (TRAIL)-mediated apoptosis in gastric epithelial cells**

TNF-related apoptosis-inducing ligand (TRAIL; also called Apo2L) is a novel TNF superfamily member with strong

homology with FasL that is capable of inducing apoptosis in a variety of transformed cell lines *in vitro*,^{22,23} but usually not in normal primary cells. T cells can kill target cells via TRAIL/TRAIL-receptor interaction,^{24,25} indicating that TRAIL might serve as a cytotoxic effector molecule in activated T cells *in vivo*. These findings suggested that TRAIL/TRAIL-receptor interaction is involved in the interaction between infiltrating T cells and gastric epithelium during *H. pylori* gastritis. Recent studies demonstrated that human gastric epithelial cells sensitized to *H. pylori* confer susceptibility to TRAIL-mediated apoptosis, suggesting a role for immune-mediated apoptosis in gastric epithelial cells by infiltrating T cells during *Helicobacter* infection.^{6,14,26} The induction of TRAIL sensitivity by *H. pylori* is dependent upon bacterial cell contact, irrespective of expression of *H. pylori* virulent factors.⁶

TRAIL-induced apoptosis occurs through a caspase signaling cascade, and resistance to TRAIL is controlled by intracellular regulators of apoptosis. Crosslinking of the TRAIL receptors leads to the formation of a death-inducing signaling complex (DISC)²⁷ assembled by the death-adaptor protein Fas-associated death-domain protein (FADD) and caspase-8 and -10.^{28–30} Activated caspase-8 is generated after DISC formation, and initiates apoptosis-inducing caspase signaling cascade. Previous studies demonstrated that TRAIL-induced apoptosis could be enhanced by *Helicobacter* and hepatitis C virus core proteins^{6,31}; however, the mechanisms leading to microbe-induced TRAIL sensitivity remain unclear. *H. pylori*-induced sensitivity to TRAIL-mediated apoptosis in gastric epithelial cells is dependent upon activation of the caspase-8 downstream pathway for conveyance of death signals to mitochondria, resulting in activation of mitochondrial pathways and removing resistance to apoptosis. *H. pylori* also induces sensitivity to TRAIL-mediated apoptosis by regulation of FLICE-inhibitory protein (FLIP) and assembly of DISC, which initiates caspase activation²⁶ and promotes apoptosis, thereby providing insight into the pathogenesis of gastric damage during *Helicobacter* infection (Figure 1). Modulation of host-cell apoptosis by bacterial interaction adds a new dimension to immune pathogenesis associated with *Helicobacter* infection.

Chemokine-mediated lymphocyte trafficking of T lymphocytes in gastric inflammation during *H. pylori* infection

During *H. pylori* infection, the degree of apoptosis induced in the stomach is affected by the inflammatory response. *H. pylori* infection induces a T cell response, as well as a number of inflammatory mediators, including cytokines and chemokines.^{7,11,14,32} These infiltrating T cells can then target and destroy gastric cells via TRAIL/TRAIL-receptor interaction. In addition to its role in inducing apoptosis by

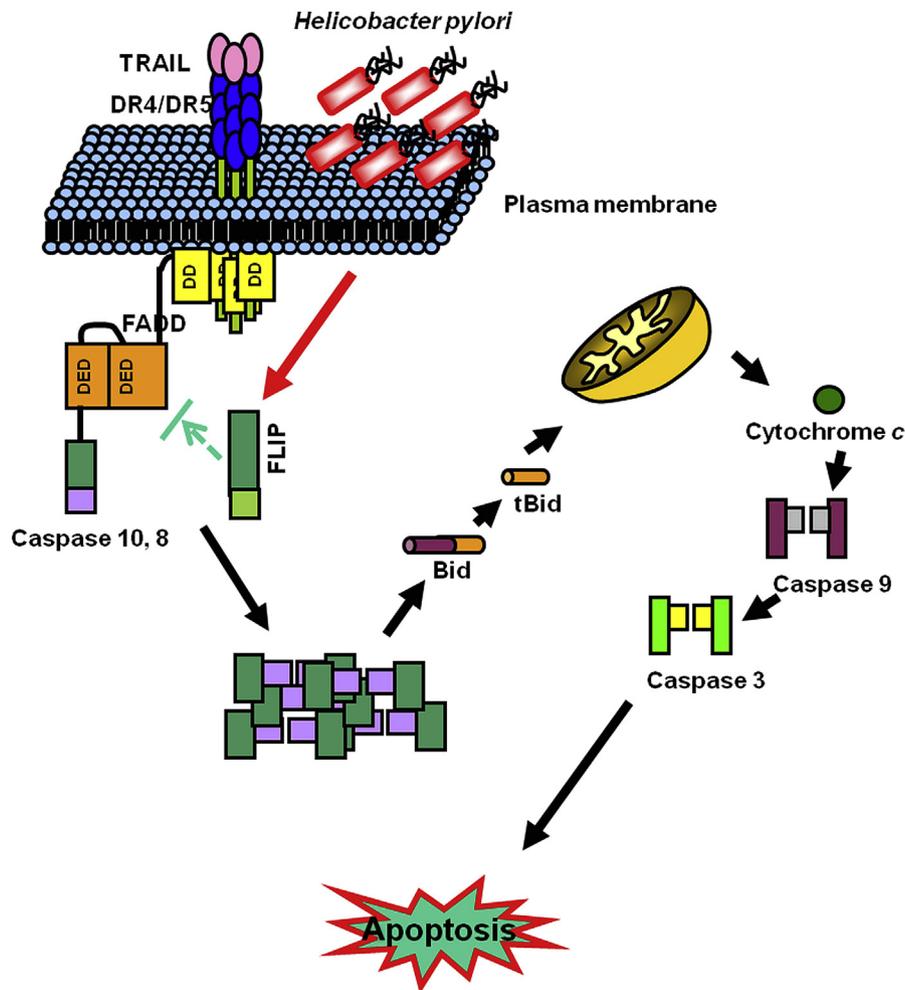


Figure 1. *Helicobacter pylori* modulates TRAIL-mediated apoptosis in gastric epithelial cells. *H. pylori* enhances assembly of TRAIL death inducing signaling complex (DISC) through downregulation of FLIP after TRAIL engagement, to activate caspase 8 cleavage, and to convey the death signal to mitochondria via cleavage of Bid, leading to activation of mitochondrial pathway and breaking the apoptosis resistance. DED = death effector domain; DD = death domain; DR4 = death receptor 4; DR5 = death receptor 5; FADD = Fas-associated death-domain protein; FLIP = FLICE-inhibitory protein; TRAIL = tumor necrosis factor-related apoptosis-inducing ligand.

binding to death receptors, TRAIL can directly stimulate T cells by binding to TRAIL receptors and transducing a reverse signal to T cells in conjunction with T cell receptor engagement, resulting in T cell proliferation and augmenting IFN- γ secretion.^{33–35} Therefore, these results support a functional role of TRAIL in *H. pylori*-induced apoptosis. Moreover, the degree of apoptosis during *H. pylori* infection may also be linked to the associated inflammatory response. Therefore, in addition to *H. pylori* virulence, mucosal damage may also be affected by the inflammatory response induced by *H. pylori* within the gastric epithelium.

Inflammation of the gastric mucosa develops in response to host immune reaction against pathogens. The stimulation of epithelial cells by *H. pylori* contributes to neutrophil and lymphocyte recruitment. As described, the features of *H. pylori*-induced inflammatory immune response are orchestrated by sequential elaboration of proinflammatory cytokines, including interleukin (IL)-10, IFN- γ , TNF- α , and IL-1 β . Accordingly, factors involved in regulating cytokine responses may confer susceptibility to or protection against *H.*

pylori-associated diseases. These results indicate that immune reaction and inflammation mediators associated with *H. pylori* plays an important role in the pathogenesis of *H. pylori*-associated diseases. Among T cells responding to *H. pylori* infection, gastric-infiltrating T cells are mostly CD45RO⁺CD69⁺CD4⁺T cells, indicating an accumulation of activated memory CD4⁺ T cells during *Helicobacter* infection.¹⁴ Recent reports indicated that Th1 and Th17 responses were induced during *H. pylori* infection,^{11,14,32} and that levels of IFN- γ and TNF- α increased in the gastric mucosa during *H. pylori* infection, augmenting *H. pylori*-induced apoptosis.^{5,6,10} These results suggest a role for immune-mediated apoptosis in gastric epithelial cells by infiltrating T cells during *Helicobacter* infection. However, the induction of immune responses and the immunopathogenic mechanism(s) of mucosal inflammation during *H. pylori* infections remain unclear, with chemokines thought to play an important role in this process.^{36,37} Chemokines are involved in acute and chronic inflammatory processes by attracting neutrophils, monocytes, and T cells to the site of inflammation via

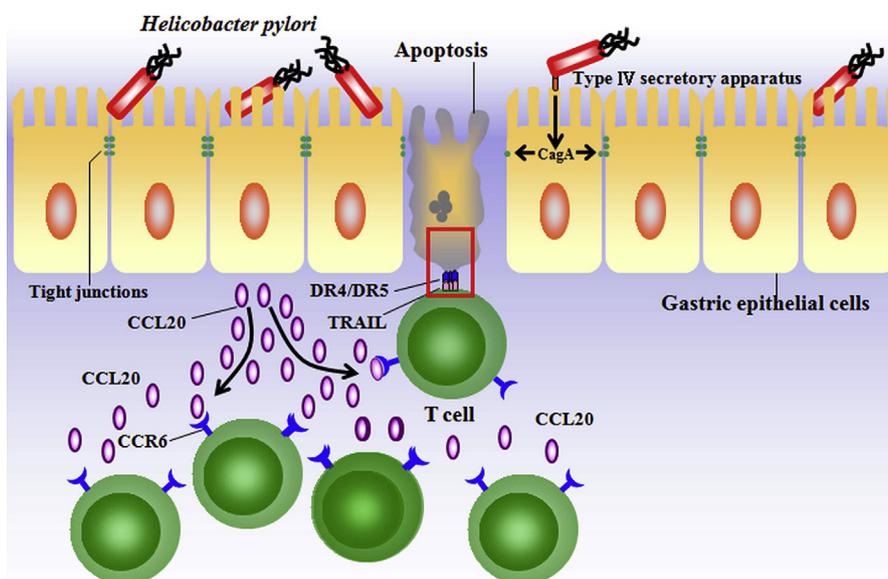


Figure 2. Immune pathogenesis of gastric mucosa damage in *Helicobacter pylori* infection. In the absence of *H. pylori* infection, there are very few infiltrating T cells recruited into the gastric mucosa. When T cells interact with human gastric epithelial cells in the presence of *H. pylori* infection, there are increased CCR6⁺ activated T cells infiltrated into gastric mucosa via CCL20 induced by *H. pylori*. The infiltrated activated T cells are recruited into gastric mucosa via CCR6, and then induce apoptosis to gastric epithelial cells via TRAIL/TRAIL receptor interaction. CagA = cytotoxin-associated gene A; DR4 = death receptor 4; DR5 = death receptor 5; TRAIL = tumor necrosis factor-related apoptosis-inducing ligand.

their corresponding chemokine receptors.^{38,39} Recent reports showed that there are specific chemokines that mediate the homing of lymphocytes in the intestines,^{37,40} suggesting that some chemokines may be involved in lymphocyte trafficking in the gut. A set of proinflammatory chemokines are also involved in *H. pylori* gastritis: Gro- α , IL-8, RANTES, IFN- γ -inducible protein-10 (IP-10; CXCL10), a monokine induced by IFN- γ (MIG, CXCL11), and CCL20 (MIP-3 α /LARC/exodus).^{14,41,42} It was demonstrated that the gastrointestinal epithelium senses invading microorganisms and produces cytokines/chemokines that attract lymphocytes and dendritic cells to the site of inflammation.⁴³ Recently, it was reported that CCR6 mediates dendritic cell localization, lymphocyte homeostasis, and immune responses in mucosal tissue.⁴⁴ CCR6, a specific β -chemokine receptor for CCL20, is selectively expressed on dendritic cells and some memory T cells,^{43,45,46} and may play a role in chemokine-mediated lymphocyte trafficking during gastric inflammation. Additionally, CCL20, the ligand of CCR6, is abundantly expressed in mouse and human inflammatory enteric mucosa,⁴⁷ and CCL20 production was upregulated in response to *H. pylori* in gastric epithelial cells when there was stimulation by the proinflammatory cytokines IL-1 β and TNF- α .^{14,48–50} These results implicated the interaction between CCL20 and CCR6 in recruiting CD45RO⁺ memory T cells to sites of inflammation in the gastric mucosa during *Helicobacter* infection (Figure 2).¹⁴

Conclusion

Human gastric epithelial cells sensitized to *H. pylori* confer susceptibility to TRAIL-mediated apoptosis, and the induction of TRAIL sensitivity by *H. pylori* is dependent upon the

activation of caspase-8 and its downstream pathway via enhanced assembly of the TRAIL DISC through down-regulation of cellular FLIP. The degree of mucosal damage is also affected by the inflammatory response induced by *H. pylori* within the gastric epithelium. These results suggest a role for immune-mediated apoptosis and mucosa damage by infiltrating T cells during *Helicobacter* infection. In conclusion, *H. pylori* enhances susceptibility of gastric epithelial cells to TRAIL-mediated apoptosis via modulation of death-receptor signaling. Modulation of host-cell apoptosis by bacterial interaction adds a new dimension to the immune pathogenesis associated with chronic *Helicobacter* infection.

Conflicts of interest

The authors declare no conflicts of interest.

Acknowledgments

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