Successful treatment of meningitis caused by multidrugresistant *Acinetobacter baumannii* with intravenous and intrathecal colistin

Yu-Huai Ho¹, Lih-Shinn Wang¹, Hui-Jen Chao², Kia-Chich Chang³, Chain-Fa Su⁴

¹Division of Infectious Diseases, Department of Internal Medicine, and Departments of ²Laboratory Medicine and ³Biotechnology, and ⁴Division of Neurosurgery, Department of Surgery, Buddhist Tzu Chi General Hospital and Tzu Chi University, Hualien, Taiwan

Received: March 15, 2007 Revised: June 15, 2007 Accepted: June 27, 2007

Multidrug-resistant *Acinetobacter baumannii* is an emergent nosocomial pathogen. A 61-year-old woman developed meningitis caused by MDRAB 27 days after receiving a surgical intervention for invasive meningioma. The patient failed to respond to high doses of meropenem and sulbactam treatment and the organism persisted in the cerebrospinal fluids for two months. The regimen was changed to intravenous and intrathecal colistin for 28 days and the patient responded well. Administration of colistin both intravenously and intrathecally could be a suitable option as a salvage therapy for meningitis due to multidrug-resistant *A. baumannii*.

Key words: Acinetobacter baumannii; Colistin; Drug resistance, multiple; Injections, spinal; Meningitis, bacterial

Introduction

Multidrug-resistant *Acinetobacter baumannii* (MDRAB) has gradually increased in importance as a nosocomial pathogen. Strains that are resistant to all available antimicrobials agents in clinical practice, including aminoglycosides, cephalosporins, fluoroquinolones, carbapenems and beta-lactamase inhibitors, are increasingly encountered. Treatment of these organisms is challenging since the choice of antibiotics is limited, and management becomes even more difficult if the clinical picture involves meningitis. We present a case of post-neurosurgical meningitis involving MDRAB that was unresponsive to carbapenem plus sulbactam, but resolved after treatment was switched to colistin.

Case Report

A 61-year-old woman was first admitted to our hospital in September 2003 with a primary complaint of

© 2007 Journal of Microbiology, Immunology and Infection

weakness in her right leg. An enormous out-bulging mass in the left parietal area was noted on computed tomography. The tumor was removed in September 2003 and the pathology assigned as invasive meningioma. In March 2006, the tumor recurred with 2 masses palpable in the scalp of the right parietal area. On the seventh hospital day, she underwent craniectomy for tumor removal and cranioplasty with brain cement. On day 11 of hospitalization, her consciousness was abruptly disturbed and follow-up computed tomography showed swelling in the left parietal lobe. Emergent craniectomy was performed to release pressure on the brain, and a dura patch was used for duraplasty. On day 27 of hospitalization, the patient received right hemicolectomy due to 2 anorectal ulcers with active bleeding despite suture ligation. On day 28 of hospitalization, external lumbar drainage was inserted to relieve the intracranial pressure. After this intervention, the patient developed fever of 38.5°C and unstable blood pressure. After ruling out other sources of infection, the artificial dura graft was suspected to be an infection focus. On day 34 of hospitalization, the artificial graft was removed and replaced by lata fascia taken from her right thigh. The external lumbar drainage was removed and an external

Corresponding author: Dr Chain-Fa Su, Division of Neurosurgery, Department of Surgery, Buddhist Tzu Chi General Hospital, 707, Section 3, Chung Yang Road, Hualien, Taiwan. E-mail: yuhuai@tzuchi.com.tw

| Variable | Day of admission | | | | | | | | |
|---------------------|--------------------|--------------|--------------------|--------------|------------------------------------|--------------|--------------|--|--|
| | 28 | 34 | 49 | 62 | 65 | 69 | 76 | | |
| CSF glucose (mg/dL) | 24 | 117 | 56 | 26 | 60 | 59 | 6 | | |
| CSF protein (mg/dL) | 345 | 39 | 28 | 158 | 109 | 65 | 112 | | |
| CSF cell count (µL) | 2910 | 210 | 20 | 440 | 82 | 200 | 243 | | |
| PMN (%) | 90 | 82 | 36 | 50 | 11 | 0 | 51 | | |
| Lym (%) | 1 | 1 | 33 | 33 | 76 | 86 | 36 | | |
| Culture | No growth | A. baumannii | A. baumannii | A. baumannii | A. baumannii | A. baumannii | A. baumannii | | |
| Antibiotics | Ox IV 2 g q6h + | | | | | | | | |
| | FI IV 2 g q8h | | | | | | | | |
| | (day 23-28) | Me 2 | g IV q8h (day 31 | -63) + | Co IV 2 million IU q8h (day 63-84) | | | | |
| | switched to | Su | 1 g IV q8h (day 39 | 9-63) | | | | | |
| | Te IVD 400 mg qd + | | | | | | | | |
| | FI IV 2 g q8h | | | | | | | | |
| | (day 28-31) | | | | | | | | |

Table 1. Summary of antibiotic treatment and laboratory data in a patient with multidrug-resistant Acinetobacter baumannii

 meningitis (day 28 to 76)

Abbreviations: CSF = cerebrospinal fluid; PMN = polymorphonuclear cells; Lym = lymphocytes; Ox = oxacillin; IV = intravenous;q6h = every 6 h; Fl = flomoxef; q8h = every 8 h; Te = teicoplanin; IVD = intravascular device; qd = once a day; Me = meropenem; Su = sulbactam; Co = colistin

ventricular drain was inserted. Three days later, cerebrospinal fluid (CSF) culture obtained during the surgery grew *A. baumannii*. The CSF data are shown in Table 1 and Table 2. The identified pathogen was resistant to all the antibiotics examined in the laboratory by disk diffusion susceptibility test, including carbapenems, cephalosporins, fluoroquinolones, aminoglycosides and aztreonam. The patient received intravenous meropenem 2 g every 8 h and sulbactam 1 g every 8 h for 30 days due to the fact that a substitution was unavailable. She had intermittent fever and CSF culture performed once per week continued to grow *A. baumannii*. All of the pathogens isolated had identical antibiograms. We started colistin 2 million IU intravenously every 8 h from day 63 of hospitalization. CSF culture continued to grow *A. baumannii* after 3 weeks of colistin administration. The external ventricular drain was revised on day 94.

Table 2. Summary of antibiotic treatment and laboratory data in a patient with multidrug-resistant Acinetobacter baumanniimeningitis (day 93-155)

| Variable | Day of admission | | | | | | | | | | |
|---------------------|--|-----------|------------|------------|------------|------------|------------------------|---------|-----------|------------|---------|
| | 93 | 97 | 99 | 100 | 101 | 104 | 118 | 121 | 125 | 128 | 155 |
| CSF glucose (mg/dL) | 4 | <0.4 | NA | NA | 31 | 60 | 60 | 64 | 58 | 51 | 54 |
| CSF protein (mg/dL) | 71 | 190 | NA | NA | 108 | 165 | 70 | 182 | 79 | 109 | 35 |
| CSF cell count (µL) | 1980 | 4230 | NA | NA | 60 | 700 | 102 | 50 | 35 | 58 | 8 |
| PMN (%) | 91 | 97 | NA | NA | 92 | 57 | 0 | 0 | 1 | 0 | 45 |
| Lym (%) | 1 | 1 | NA | NA | 6 | 27 | 92 | 73 | 92 | 66 | 39 |
| Culture | A. bau- | A. bau- | NA | NA | No | No | No | A. bau- | A. bau- | No | No |
| | mannii | mannii | | | growth | growth | growth | mannii | mannii | growth | growth |
| Antibiotics | Mel | Me IV 2 g | | Co IT | Co IT | Co IT | Co IT 60,000 IU q48h + | | | Co IT | Dis- |
| | q8h + Su IV 1 g q8h (day 84-98) | | 20,000 IU | 40,000 IU | 60,000 IU | 30,000 IU | Co IV 2 million IU q8h | | 80,000 IU | continued | |
| | | | q24h + | q24h + | q24h + | q24h + | (day 114-126) | | | q24h + | Co on |
| | | | Co IV 2 | Co IV 2 | Co IV 2 | Co IV 2 | | | | Co IV 2 | day 155 |
| | | | million IU | million IU | million IU | million IU | | | | million IU | |
| | | | q8h | q8h | q8h | q8h | | | | q8h | |
| | | | (day 99) | (day 100) | (day 101- | (day 103- | | | | (day 127- | |
| | | | | | 102) | 113) | | | | 155) | |

Abbreviations: CSF = cerebrospinal fluid; PMN = polymorphonuclear cells; Lym = lymphocytes; NA = not available; Me = meropenem; IV = intravenous; q8h = every 8 h; Su = sulbactam; Co = colistin; IT = intrathecal; q24h = every 24 h; q48h = every 48 h

On day 99 of hospitalization, intrathecal injection of colistin 20,000 IU per day was given through an external shunt. The daily dosage of intrathecal colistin was slowly increased to 60,000 IU in combination with intravenous colistin 2 million IU every 8 h. CSF culture was sterile on the second day of intrathecal therapy and remained sterile for the next 3 weeks. Nevertheless, A. baumannii reappeared when we reduced the dose of intrathecal colistin to 60,000 IU every 48 h. The intrathecal dosage was elevated to 80,000 IU every 24 h from day 127 of hospitalization. CSF culture was sterile on the next day and we removed the ventriculoperitoneal shunt that was concomitant with external ventricular drainage on day 135. A permanent ventriculoperitoneal shunt was inserted on day 155 after the CSF culture had been confirmed as remaining sterile for 4 weeks. The patient has been followed up for 6 months, with no evidence of relapse.

Discussion

A. baumannii is a Gram-negative, non-fermenting coccobacillus that is widespread in nature. It does not have fastidious growth requirements and is able to grow at various temperature and pH conditions [1]. A. baumannii survives in either moist or dry conditions in the hospital environment and has intrinsic resistance to many antimicrobial agents. These factors have contributed to the emergence of A. baumannii as a leading pathogens in nosocomial infections.

The presence of carbapenem-resistant *A. baumannii* was first reported in the United States in 1991 [2]. Since then, carbapenem-resistant *A. baumannii* infections and hospital outbreaks have been reported in many countries. Furthermore, a so-called MDRAB was first reported in Taiwan in 1998 [3]. MDRAB is resistant to almost all commercially available antibiotics, including carbapenems, cephalosporins, aztreonam, aminoglycosides and fluoroquinolones. MDRAB was first noticed in our hospital in 2002. We used combination therapy of carbapenem (imipenem or meropenem) with sulbactam, and the clinical response was fairly satisfactory. None-theless, the consequences can be grave when the infection involves meningitis.

Colistin is one of the polymyxin antibiotics produced by *Bacillus colistinus*. Polymyxins were discovered in 1947 and have been available since 1959 for the treatment of infections caused by Gram-negative bacteria [4,5]. However, early clinical reports noted a high incidence of nephrotoxicity and neurotoxicity



Fig. 1. Molecular typing of the cerebrospinal fluid Acinetobacter baumanii isolate by pulsed-field gel electrophoresis

associated with colistin. Colistin was thus abandoned and replaced by other less toxic antibiotics, such as beta-lactams. Nowadays, the emergence of multidrugresistant Gram-negative pathogens has led to a resurgence in the use of polymyxin. The cationic polypeptide of colistin interacts with anionic lipopolysaccharide molecules in the outer membrane of Gram-negative bacteria, which leads to derangement of the cell membrane [6]. Polymyxins provide bacteriostatic activity at low concentrations, bactericidal activity at high concentrations and potent antiendotoxic activity [7,8].

At present, clinical articles on the intrathecal use of colistin are scarce. Variable doses of intrathecal colistin, ranging from 1.6 to 20 mg, have been used in different studies [9-13]. However, intrathecal administration of colistin 60,000 IU every 48 h failed to eradicate MDRAB in this case. Molecular typing by pulsed-field gel electrophoresis showed that serial CSF isolates were of the same clonal type (Fig. 1). Increasing the intrathecal colistin dose to 80,000 IU every 24 h and combining this with intravenous colistin 2 million IU every 8 h saw eventual resolution of the infection.

Generally, intrathecal or intraventricular administration of colistin has been well tolerated, except in 1 study, which reported development of numbness on the patient's left arm. The symptom subsided after the dose of intrathecal colistin was reduced [14].

From the case presented here, we propose that cases of meningitis caused by MDRAB could be treated with intrathecal colistin 80,000 IU per day in combination with intravenous colistin 2 million IU every 8 h. The combination therapy could be an effective alternative treatment.

References

- Bergogne-Bérézin E, Towner KJ. *Acinetobacter* spp. as nosocomial pathogens: microbiological, clinical and epidemiological features. Clin Microbiol Rev. 1996;9:148-65.
- Go ES, Urban C, Burns J, Kreiswirth B, Eisner W, Mariano N, et al. Clinical and molecular epidemiology of *Acinetobacter* infections sensitive only to polymyxin B and sulbactam. Lancet. 1994;344:1329-32.
- Kuo LC, Teng LJ, Yu CJ, Ho SW, Hsueh PR. Dissemination of a clone of unusual phenotype of pandrug-resistant *Acinetobacter baumannii* at a university hospital in Taiwan. J Clin Microbiol. 2004;42:1759-63.
- Benedict RG, Langlykke AF. Antibiotic activity of *Bacillus* polymyxa. J Bacteriol. 1947;54:24-5.
- Ross S, Puig JR, Zaremba EA. Colistin: some preliminary laboratory and clinical observations in specific gastroenteritis in infants and children. Antibiot Annu. 1959-1960;7: 89-100.
- 6. Falagas ME, Kasiakou SK. Colistin: the revival of polymyxins for the management of multidrug-resistant gram-negative bacterial infections. Clin Infect Dis. 2005;40:1333-41.
- 7. Horton J, Pankey GA. Polymyxin B, colistin, and sodium colistimethate. Med Clin North Am. 1982;66:135-42.

- 8. Evans ME, Feola DJ, Rapp RP. Polymyxin B sulfate and colistin: old antibiotics for emerging multiresistant gram-negative bacteria. Ann Pharmacother. 1999;33:960-7.
- Katragkou A, Roilides E. Successful treatment of multidrugresistant *Acinetobacter baumannii* central nervous system infections with colistin. J Clin Microbiol. 2005;43:4916-7.
- Benifla M, Zucker G, Cohen A, Alkan M. Successful treatment of *Acinetobacter meningitis* with intrathecal polymyxin E. J Antimicrob Chemother. 2004;54:290-2.
- Fernandez-Viladrich P, Corbella X, Corral L, Tubau F, Mateu A. Successful treatment of ventriculitis due to carbapenemresistant *Acinetobacter baumannii* with intraventricular colistin sulfomethate sodium. Clin Infect Dis. 1999;28:916-7.
- Vasen W, Desmery P, Ilutovich S, Di Martino A. Intrathecal use of colistin. J Clin Microbiol. 2000;38:3523.
- Kasiakou SK, Rafailidis PI, Liaropoulos K, Falagas ME. Cure of post-traumatic recurrent multiresistant Gram-negative rod meningitis with intraventricular colistin. J Infect. 2005;50: 348-52.
- Schina M, Spyridi E, Daoudakis M, Mertzanos E, Korfias S. Successful treatment of multidrug-resistant *Pseudomonas aeruginosa* meningitis with intravenous and intrathecal colistin. Int J Infect Dis. 2006;10:178-9.